

DIAGNOVITAL

SARS-CoV-2 E484K Mutation Detection Kit

Qualitative RT-PCR-based detection of SARS-CoV-2 E484K Mutation

For Research use only. Not for use in diagnostic procedures.

RUO

REF



090R03025 25 tests

090R03050 50 tests

090R03100 100 tests

Table of Contents

Intended Use	3
Product Description	3
Materials Provided	4
Additional Materials Required	4
Storage	4
Performance Characteristics	5
Considerations Before Starting	5
Sample Preparation	6
Reaction Setup	6
Analysis & Troubleshooting	9
Limitations	12
Trademarks	12
Symbols	13

Intended Use

DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit is a real-time RT-PCR-based test for the detection and differentiation of SARS-CoV-2 E484K mutation in respiratory specimens. **This kit is designed to run only samples that have previously proven to be SARS-CoV-2 positive.**

DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit detects E484K mutation in extracted SARS-CoV-2 RNA from nasopharyngeal and oropharyngeal swab samples during infection. Positive results indicate the presence of SARS-CoV-2 RNA and allow differentiation between E484 and K484 in the Spike protein. Spike protein of wild type SARS-CoV-2 has glutamic acid (E) amino acid at 484th location, which is defined as E484. The mutant SARS-CoV-2 E484K strain has lysine (K) amino acid at 484th location in Spike protein and it is defined as K484. The E484K is a mutation which occurs in different lineages and has already been found in the South African (B.1.351) and Brazilian (B.1.1.28) lineage. The mutation is in the spike protein and appears to have an impact on the body's immune response and, possibly, vaccine efficacy. The P.1. lineage described in Japan and Manaus, the P.2 lineage (also known as B.1.1.248 lineage, Brazil) and 501.V2 (South Africa) exhibit this mutation. A limited number of B.1.1.7 genomes with E484K mutation have also been detected. Monoclonal and serum-derived antibodies are reported to be from 10 to 60 times less effective in neutralizing virus bearing the E484K mutation.

Product Description

DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit is a real-time RT-PCR-based detection and differentiation system for SARS-CoV-2 E484K mutation. **This kit is designed to run only on samples that have previously proven to be SARS-CoV-2 positive.**

DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit also detects the presence of a sequence specific human target (RNaseP) serving as a human extraction control (HEC) in addition to the wild type (E484) and the mutant (K484) sequences in different channels.

REAL TIME PCR-BASED DETECTION OF SARS-CoV-2

The first step in the detection of SARS-CoV-2 E484K mutation is the conversion of viral RNA into cDNA. Afterwards, the viral target sequences and the RNaseP (HEC) are simultaneously amplified in one reaction with amplification monitored in real time through the use of fluorescently labelled probes: upon incorporation into the newly amplified DNA strands, the fluorophore is released and an increase in fluorescence signal can be observed.

With **DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit** differentiation between the viral targets is achieved through the use of two different fluorophores that are detected in three different channels: FAM™ for wild type SARS-CoV-2 E484, HEX/VIC for K484 mutation and the RNaseP (HEC) is detected in the Cy5 channel.

Due to the intrinsic mutation rate of viruses, it is possible that mutations in the target sequence occur and accumulate over time. This can lead to false-negative results with a PCR-based detection approach.

DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit is validated with Applied Biosystems Quant Studio 5 Dx and compatible with BioRad CFX96, Applied Biosystems Quant Studio 5 Real-Time PCR Systems calibrated FAM™, HEX/VIC and Cy5 channels.



RTA Laboratuvarları

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Revision Date/ Revision No: -/-



Materials Provided

Table 1: DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit contents

	Reagents	Quantity and Volume (25 tests)	Quantity and Volume (50 tests)	Quantity and Volume (100 tests)
1	PCR Master Mix	1 × 375 µl	1 × 750 µl	1 × 1500 µl
2	Positive Control	1 × 38 µl	1 × 75 µl	1 × 150 µl
3	Nuclease-free dH ₂ O	1 × 38 µl	1 × 75 µl	1 × 150 µl



IMPORTANT! The table above reflects the standard kit color scheme. Due to supplier issues during the COVID-19 crisis, individual tube cap colors may be substituted due to availability. Always check the labeling of the reagent prior to use.

Additional Materials Required

- Suitable means & equipments for nucleic acid extraction
- Real-time PCR detection system equipped for FAM™, HEX/VIC and Cy5 detection
- Adjustable pipettes & fitting filtered pipette tips
- Appropriate personal protective equipment & workspaces for working with potentially infectious samples
- Surface decontaminants such as DNAzap™ (Life Technologies), DNA Away™ (Fisher Scientific), RNAse Away™ (Fisher Scientific), 10% bleach (1:10 dilution of commercial 5.25-6.0% sodium hypochlorite)
- Nuclease-free tubes / strips / plates to prepare dilutions, master mixes etc.
- Real-Time PCR reaction tubes/plates/capillaries

For **BIO-RAD CFX96**: Hard-Shell Thin-Wall 96-Well Skirted PCR Plates with Bar Codes (BIO-RAD, Cat#: HSP-9955), Hard-Shell Thin-Wall 96-Well Skirted PCR Plates (BIO-RAD, Cat#: HSP-9655), Microseal 'B' Adhesive Seals, optically clear (BIO-RAD, Cat#: MSB-1001), Individual PCR Tubes, Low Tube Strips, 8-tubes strip, 0.2 ml Low Profile, White (BIO-RAD, Cat#: TLS0851), Flat Cap Strips, Optically Clear, 8-cap strip, 0.2 ml (BIO-RAD, Cat#: TCS0803).

For **Applied Biosystems Quant Studio 5 Dx- Quant Studio 5 Real-Time PCR System**, MicroAmp® Optical 96-Well Reaction Plate (Thermo Fisher, Cat#: 4306737), MicroAmp® Optical Adhesive Film (Thermo Fisher, Cat#: 4311971), MicroAmp® Optical 8-Tube Strip, 0.2 mL (Thermo Fisher, Cat#: 4316567), MicroAmp® Optical 8-Cap Strips (Thermo Fisher, Cat#: 4323032)

- Suitable storage options for reagents and specimen (4°C, -20°C, -70°C)

Storage

- Store all components at -15°C /-25°C and avoid more than 3 freeze and thaw cycles.
- Protect the qPCR master mix from light as prolonged exposure can diminish the performance of the fluorophores.
- If the kit components have been damaged during transport, contact RTA Laboratories. Do not use as performance may be compromised.
- Keep reagents separate from sample material to avoid contamination.
- Do not use after the designated expiry date.



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Performance Characteristics

Diagnostic specificity

A total of 192 clinical samples that were collected from patients with Covid symptoms were analyzed by DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit and next generation sequencing analysis for first line testing to determine the positive samples and 10 of them were found to be E484K mutation positive and 182 of them were mutation negative. The positive percent agreement (PPA) of DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit with respect to NGS is 100% and the negative percent agreement (NPA) is 100%. All of the Internal Controls (RNaseP) have been tested positive.

Table 2. DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit –NGS qPCR correlation study

NP swab		Comparator (NGS and qPCR)	
		Mutant	Wild Type
DIAGNOVITAL SARS-CoV-2 E484K Mutation Detection Kit	K484	10	0
	E484	0	182
	Total	10	182

Considerations Before Starting

Biosafety

- Wear appropriate personal protective equipment (e.g., gowns, powder-free gloves, eye protection) when working with clinical specimens.
- Specimen processing should be performed in a certified class II biological safety cabinet following biosafety level 2 or higher guidelines.
- For more information, refer to:
- Interim Guidelines for Collecting, Handling, and Testing Clinical Specimens from Patients Under Investigation (PUIs) for 2019 Novel Coronavirus (SARS-COV-2)
<https://www.cdc.gov/coronavirus/2019-ncov/lab/guidelines-clinical-specimens.html>
- Biosafety in Microbiological and Biomedical Laboratories 6th edition available at <http://www.cdc.gov/biosafety/publications/>.
- The use of **DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit** and data evaluation is restricted to trained laboratory personnel only.
- Good laboratory practice is essential for optimal performance of this assay. Special care must be taken avoid contamination of the components of the kit. All reagents must be closely monitored for impurities and contamination. Discard suspicious reagents according to local guidelines and regulations.

Specimens

Only use appropriate specimens for testing, such as:

- Respiratory specimens including nasopharyngeal / oropharyngeal swabs and bronchoalveolar lavage.
- Swab specimens should be collected only on swabs with a synthetic tip (such as polyester or Dacron®) with plastic shafts. Swabs with calcium alginate or cotton tips with wooden shafts are not acceptable.



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Specimens - Handling and Storage

- Specimens can be stored at 2-8°C for up to 72 hours after collection.
- If a delay in extraction process is expected, store specimens at -20°C or lower.
- Extracted nucleic acids should be stored at -20°C or lower.

Do not use specimens if

- they were not kept at 2-8°C (≤ 4 days) or frozen at -20°C or lower.
- they are insufficiently labelled or lack documentation.
- they are not suitable for this purpose (see above for suitable sample material).
- the specimen volume is insufficient.

Sample Preparation

- The performance of RT-PCR assays strongly depends on the amount and quality of sample template RNA. It is strongly recommended to qualify and validate RNA extraction procedures for recovery and purity before testing specimens.
- **DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit** is validated with RTA Viral RNA Isolation Kit.
- **DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit** is compatible with Tianlong Generotex96 Extraction System and QIAamp® MinElute Virus Spin Kit, VERSANT® Sample Preparation 1.0 Reagents.
- Validated Real Time PCR systems: Applied Biosystems Quant Studio 5 Dx.
- Compatible Real Time PCR systems: BioRad CFX96, Applied Biosystems Quant Studio 5 Real-Time PCR Systems.
- Store and keep residual specimens and extracted nucleic acids at -20°C or -80°C.
- Only thaw the number of specimen extracts that will be tested in a single day.
- Do not freeze/thaw extract more than once before testing as each freeze/thaw cycle will decrease the RNA quality.
- It may be possible to use patient samples directly, depending on the sample type. However, this may require a prior lysis step and titration of the amount on sample that can be used without inhibiting the reaction. This procedure has not been validated, use of isolated RNA is recommended.

Reaction Setup

1. Make sure that all necessary equipment and devices are suitable, calibrated and functional before starting the experiments.
2. Decontaminate equipment and workspace and prepare everything needed for the following experiment to keep the workflow short and repeatable.
3. Switch on the PCR detection system and program it to avoid delays after setting up the reactions.
4. Thaw all components of **DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit** on ice and mix gently but thoroughly to ensure even distribution of components. Collect liquid at the bottom of the tube with a quick spin (via microcentrifuge).



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5. The PCR Master Mix provided with DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit is ready to use. One reaction will be prepared for each sample. A separate reaction should be prepared for Negative Control (NTC) and Positive Control (TPC).

Component	Volume (µl)
PCR Master Mix	15
RNA Isolate/ TPC/ NTC	5
Total	20

6. Distribute **15 µl** PCR Master Mix to your strips/plate and add **5 µl your samples**. (An example setup is given in Fig1).

	1	2	3	4	5	6	7	8	9	10	11	12
A	S1	S9	S17	S25	S33	S41	S49	S57	S65	S73	S81	TPC
B	S2	S10	S18	S26	S34	S42	S50	S58	S66	S74	S82	S89
C	S3	S11	S19	S27	S35	S43	S51	S59	S67	S75	S83	S90
D	S4	S12	S20	S28	S36	S44	S52	S60	S68	S76	S84	S91
E	S5	S13	S21	S29	S37	S45	S53	S61	S69	S77	S85	S92
F	S6	S14	S22	S30	S38	S46	S54	S62	S70	S78	S86	S93
G	S7	S15	S23	S31	S39	S47	S55	S63	S71	S79	S87	S94
H	S8	S16	S24	S32	S40	S48	S56	S64	S72	S80	S88	NTC

Figure 1: Example pipetting scheme for the distribution of master mixes with the individual assay mixes

7. Transfer the reactions to the PCR device, then proceed according to these guidelines:

Table 3: DIAGNOVITAL SARS-CoV-2 E484K Mutation Detection Kit Thermal Profile

Step	Cycles	Temperature	Duration
Reverse Transcription	1	50°C	10 minutes
Initial Denaturation	1	95°C	2 minutes
Amplification	40	95°C	5 seconds
		57°C* 	30 seconds

*Enable Data Collection for **FAM™** (Wild Type E484 SARS-CoV-2), **HEX/VIC** (K484 SARS-CoV-2 Mutation) and **Cy5** (HEC).

8. Once the run is finished, do not open the reaction tubes to avoid contamination and discard according to local guidelines and regulations. Do not autoclave as this may contaminate laboratory equipment with amplicons.



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Analysis & Troubleshooting

EXEMPLAR RESULTS

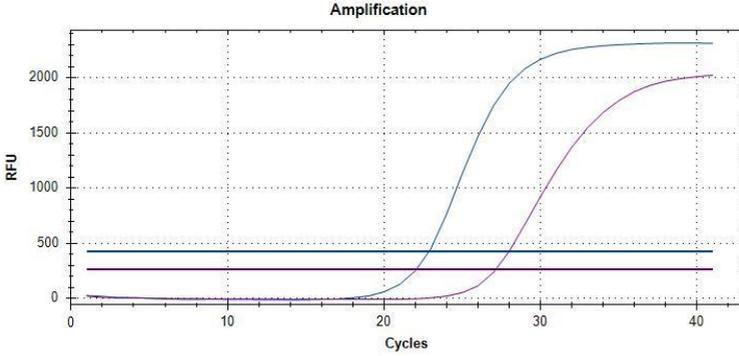


Figure 2: Blue Curves: E484 Wild Type sample at the FAM channel, Purple Curves: internal control at the Cy5 channel.

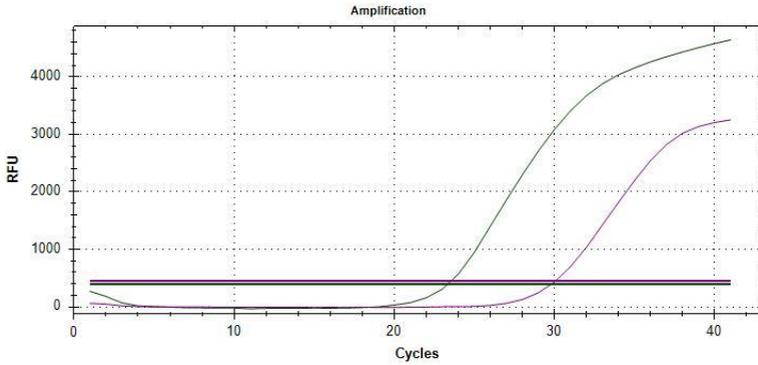


Figure 3: Green Curves: K484 Mutant positive sample at the HEX channel, Purple Curves: internal control at the Cy5 channel.

- **dH₂O controls (NTC) must not give a positive Ct for any assay.** If they do, the reaction was contaminated with sample RNA / DNA. Decontaminate equipment and workspace and repeat the reactions.
- **For a sample to be considered negative for E484K mutation, the FAM™ channel must give a positive Ct value.** Amplification of the HEC in Cy5 channel is expected around Ct 20-38. Should the HEC fail to amplify, the sample must still be considered positive. This outcome is possible when having an unusually high virus titer, or the sample was not of human origin, but cell culture derived or analysis of surface contamination.

- For a sample to be considered **positive for E484K mutation**, the **HEX/VIC** channel must give a **positive Ct value**.
- All reactions containing RNA isolate must give **positive Ct values for the internal control assay**. The Ct values should be **< 38 cycles**. Failure to amplify the internal control indicates a flawed RNA extraction or loss of RNA isolate due to RNase contamination. The sample is not sufficient, results cannot be interpreted.
- For the positive control, a **positive Ct at the FAM and HEX/VIC channels must be observed**. The Ct value for the positive control should be **20±3**. If the Ct value does not correspond to the expected value or the **positive control** was not tested positive, PCR was compromised. Check the reaction setup and PCR device settings and repeat the reactions. Repeated freeze and thaw cycles of the **positive control** can compromise its quality resulting in late Ct values.
- If Ct is <38 for each of the FAM, HEX/VIC and Cy5 channels, the result in the respective channel should be considered as **POSITIVE**, if Ct is > 38 or no value is received, the result in the relevant channel should be considered as **NEGATIVE**.

FAM	HEX/VIC	Cy5	Result
-	-	+	The sample is negative for SARS-CoV-2. This kit is designed to run only on samples that have previously proven to be SARS-CoV-2 positive. -Be sure that the sample is SARS-CoV-2 positive. -RNA may be degraded during transport, extraction or storage. -Variations in the probe binding regions-sequence analysis with a different method is recommended.
+	-	+	The sample is negative for E484K mutation.
-	+	+	The sample is positive for E484K mutation.
-	-	-	No amplification in any channel indicates flawed RNA isolation, sample degradation or PCR inhibition. Results cannot be interpreted.
+	+	-	Expected result for the Positive Control (TPC) .
-	-	-	Expected result for the Negative Control (NTC) .

Device Setup

For **QUANTSTUDIO5DX**, In the Home screen, create or open a template. In the New Experiment pane, click on Create New Experiment button to create a new template. In the Properties tab, enter the template information. In the Method tab, adjust the reaction volume and set up appropriate thermal profile. In the Plate tab (Quick Setup), assign plate attributes by selecting the Passive Reference from the dropdown list. In the Plate tab (Quick Setup), define and assign well attributes and select wells in the Plate Layout or the Well Table. Then assign samples and targets to selected wells.

Note: New sample or target names entered in the Quick Setup subtab are automatically populated with default values for Reporter as FAM, for Quencher as NFQ-MGB and for Task as Unknown. Edit these values in the Advanced Setup subtab. For TaqMan probes, NFQ-MGB option must be used as the quencher. Then start the run.

For **BIORAD CFX 96**, in the Software application, open the protocol from the File menu item. Create the appropriate protocol for the kit intended to be used. In the Plate tab, define and assign well attributes and select wells in the Plate Layout or the Well Table. Then assign samples and targets to selected wells. Then start the run.



Baseline and Threshold Settings

After the run,

For **QUANTSTUDIOSDX**, click on Show Plot Settings button to change graph type from log scale to linear scale. The target can be changed in Target section. Then click on the Analysis Setting button to adjust the baseline threshold. Unlick the Automatic Threshold and Unlick the Automatic Baseline. Set the baseline start cycle to 7-8 and baseline end cycle to 20 in order to normalize the graphics.

For **BIORAD CFX 96**, threshold can be adjust according to the the ratio of FAM vs HEX signal height.

The amplification curve baseline is one of the parameters that can affect PCR results. In case the baseline is incorrectly set, a Ct value can be displayed even if no real amplification occurred. Auto threshold is used with **DIAGNOVITAL® SARS-CoV-2 E484K Mutation Detection Kit** for PCR Detection systems. If the increase of a sample in any channel is **less than 10%** of the increase of the highest increasing sample in the same channel, this increase is considered as **NEGATIVE**. In some cases, the threshold should be set manually to avoid background fluorescence. For each sample, the ratio of FAM vs HEX signal height should be checked; the signal that outweighs the other by 3-fold or more should be regarded as positive, since only one of them should amplify.

Troubleshooting

PROBLEM	POTENTIAL REASONS	SOLUTION
Negative Result for Internal Control	PCR Master Mix may not have been homogenous.	Pipetting should be performed for PCR Master Mix.
	RNA isolation may not be performed properly.	The study should be repeated from isolation.
	Isolate may include inhibitor.	Real Time PCR stage should be repeated by diluting the isolate 1/10.
Positive Result for NTC	Contamination may have occurred.	Contamination may have occurred from the work area to the consumable items being worked on. It is recommended to dispose of consumables and open new ones and clean the environment first with 10% NaClO solution and then with 70% Alcohol.
Samples with Ct > 38	Nonspecific amplification.	The study should be repeated. If the same result is obtained, the sample is considered negative.



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Limitations

- This kit is designed to run only samples that have previously proven to be SARS-CoV-2 positive.
- For reliable results, it is essential to adhere to the guidelines given in this manual. Changes in reaction setup or cycling protocol may lead to failed experiments.
- Depending on the sample matrix, inhibitors may be present in the isolated RNA and disable reverse transcription and/or PCR amplification. If this is the case, another sample type or isolation method may be beneficial.
- Spontaneous mutations within the target sequence may result in failure to detect the target sequence.
- Results must always be interpreted in consideration of all other data gathered from a sample. Interpretation must be performed by personnel trained and experienced with this kind of experiment.

Trademarks

NucliSens® (bioMérieux), QIAamp®, RNeasy® (QIAGEN), ChargeSwitch® (Invitrogen), FAM™ (Life Technologies), DNAZap™, DNA Away™, RNase Away™

Registered names, trademarks, etc. used in this document, even if not specifically marked as such, are not to be considered unprotected by law



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Symbols



Expiry Date



Lot/ Batch



Catalog Number



Temperature Limitation



Caution



Manufacturer



Consult instructions for use or
consult electronic instructions for use



Contains sufficient for (n) amount tests



Research Use Only

RTA LABORATUVARLARI BİYOLOJİK ÜRÜNLER İLAÇ VE MAKİNE SAN. TİC. A.Ş.

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